

Instructions for Labeling Proteins with Europium

Reagents. Store all reagents at 4 C.

- Reagent 1** Proprietary chelate (7.3×10^{-6} mol)
Reagent 2 Reconstitution solution (2 ml).
Reagent 3 1 M EuCl_3 (0.2 ml)
Reagent 3 0.2 M carbonate buffer (pH 8.9)

Generic conjugation procedure

1.1 Protein preparation.

Transfer protein to a suitable solution free of nucleophiles, such as amine and sulfhydryl groups. We recommend that the protein conjugation be done in 0.2 M carbonate buffer pH 8.5 to 8.9. If possible, start by dissolving the solid protein in the reaction buffer.

1.2 Reconstitution of proprietary chelate and chelation of europium.

To the container of Reagent 1, add in sequence 393 μl of Reagent 2 and 6.66 μl (6.66×10^{-6} mol) of Reagent 3. Cap container and vortex approximately 2 minutes until solid dissolves. Let the resulting solution sit for 5 minutes.

1.3 Protein conjugation.

At room temperature and with gentle mixing of your protein, add approximately one, two, three or more mols of chelate per mol of protein. Continue mixing for 2 hours.

1.4 Purification.

Use a suitable size separation procedure for purification of your protein, such as gel filtration or dialysis against a suitable buffer for your protein.

Example preparation

2.1 Protein preparation.

Dissolve 50 mg of albumin (7.3×10^{-7} mol) in 10 ml of Reagent 4.

2.2 Reconstitution of proprietary chelate and chelation of europium.

To the container of Reagent 1, add in sequence 393 μl of Reagent 2 and 6.66 μl (6.66×10^{-6} mol) of Reagent 3. Cap container and vortex approximately 2 minutes until solid dissolves. Let the resulting solution sit for 5 minutes.

2.3 Protein conjugation.

At room temperature and with gentle mixing of the albumin, add all of the reconstituted chelate (7.3×10^{-6} mol) to the albumin. Continue mixing for 2 hours.

2.4 Purification.

Dialyze the conjugated albumin against a 10K MWCO membrane. Store at 4°C.

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